

# Imaging and Image Processing for Plant Phenotyping

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# IBG-2: Plant Sciences

*Dynamic plants in a dynamic environment*

- Founded in 2001
- 160 employees
- Main topics:  
Plant Phenotyping  
Sustainable Bio-economy
- Multi-disciplinary: biology, chemistry, physics, mathematics, computer sciences, engineering



# Why Plants?

- Plants are socially and industrially relevant
  - 4F: Food, Feed, Fibre, Fuels, but also
  - bio-chemicals like in medical plants etc.
- Industries: Farming, Seed production, plant production, fertilizers



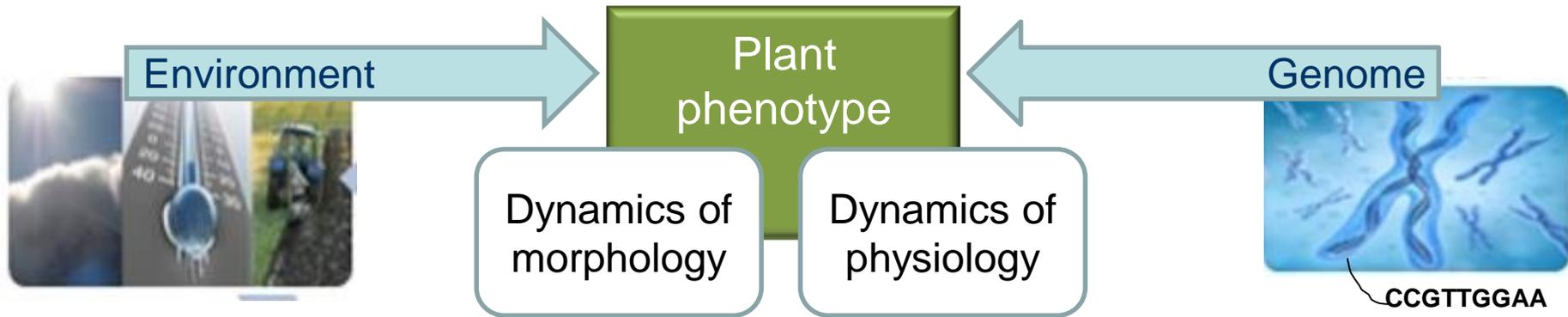
## **Central Question**

How to grow plants optimally under natural or controllable conditions?



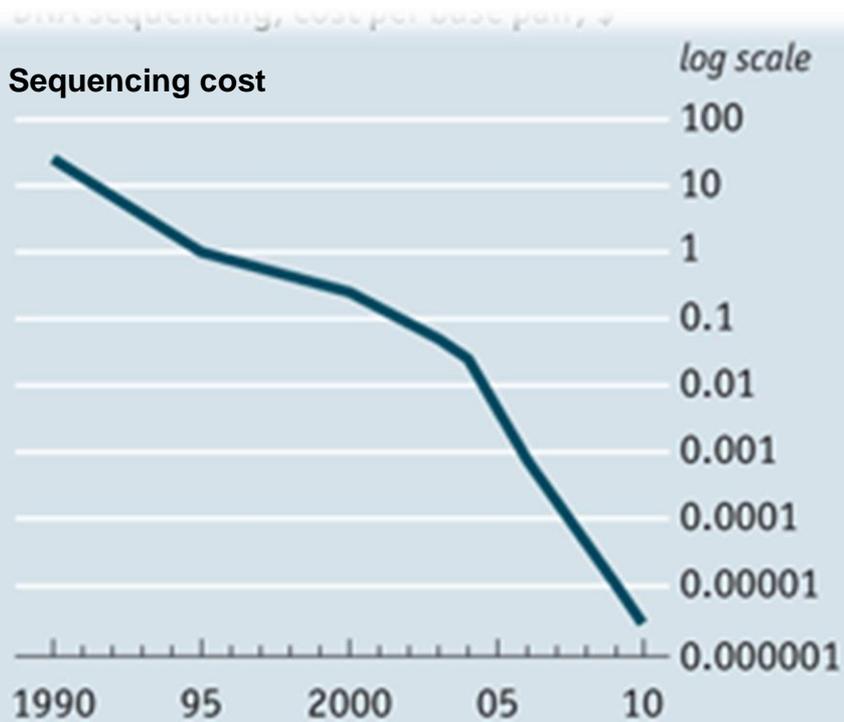
Mechanistic understanding of interactions between environmental conditions and plant traits required!

# What exactly is phenotyping?



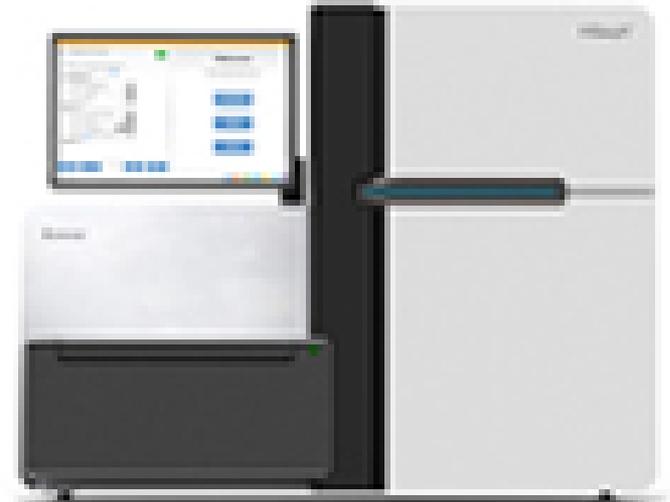
# Genotyping

GENOTYPE



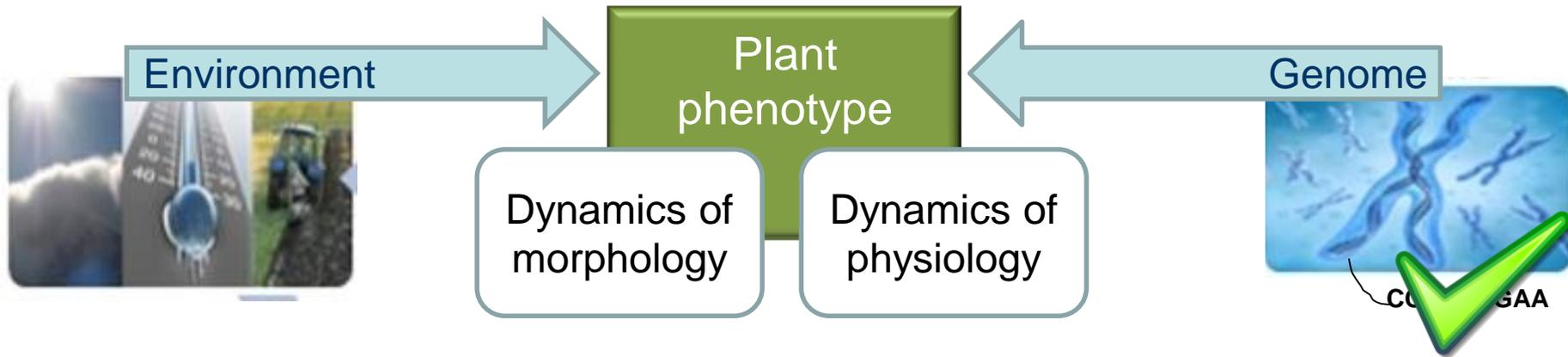
Sequencing cost is falling all the time

Illumina HiSeq X10



600 Gbases  
per day

# What exactly is phenotyping?



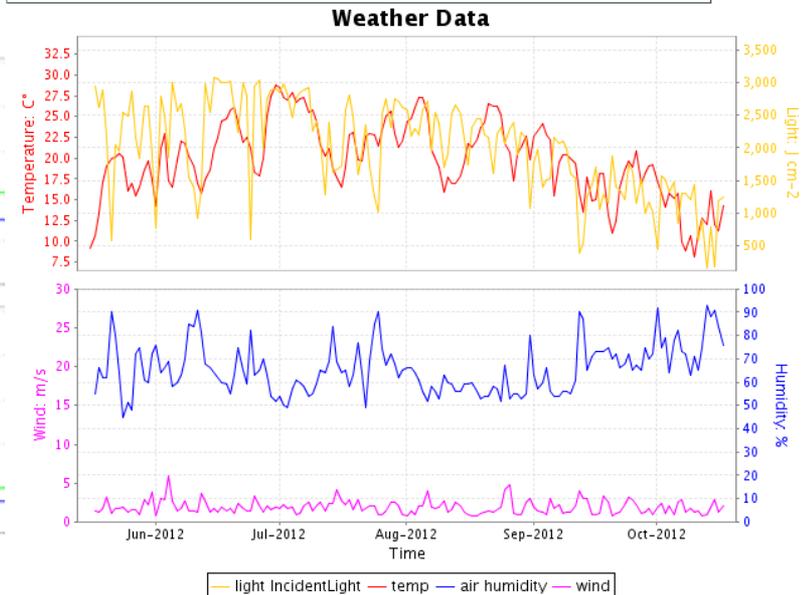
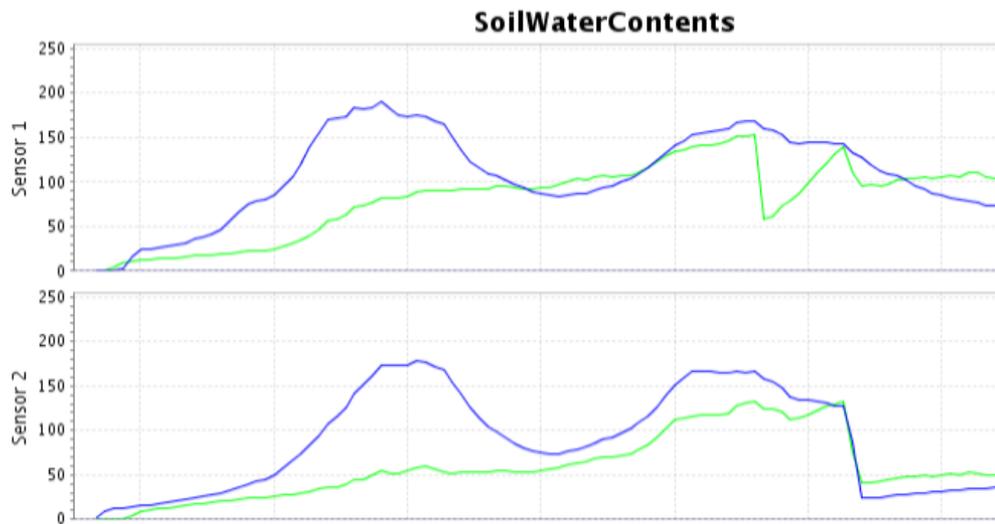
# Environmental data

## Weather data, Soil Water content

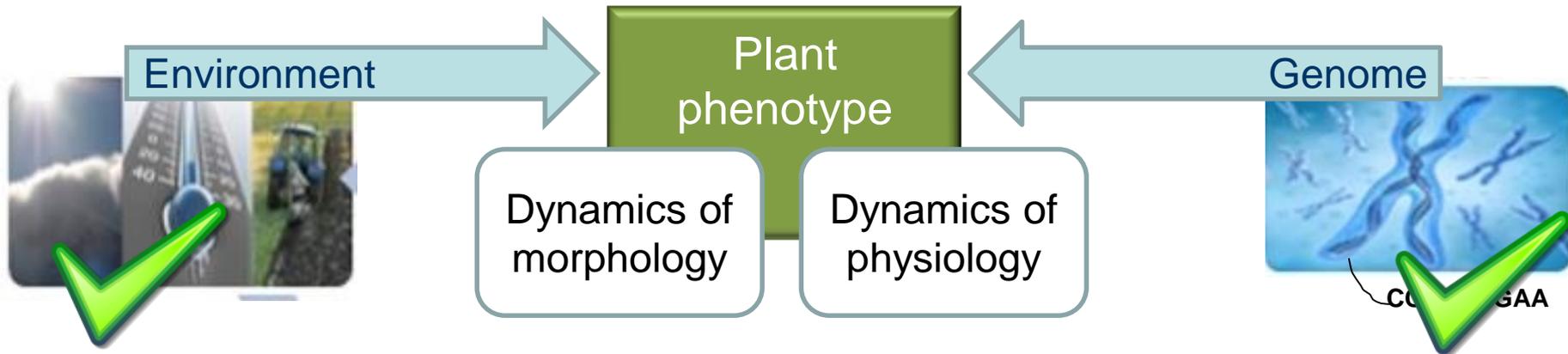
- temperature, light, wind, humidity
- Summarized by day
- Min, max, average etc
- Structure and chemical properties of soil



temperature:	DAILY_AVERAGE	light:	DAILY_MAX
wind:	DAILY_MAX	humidity:	DAILY_MIN
<input type="button" value="show"/>			



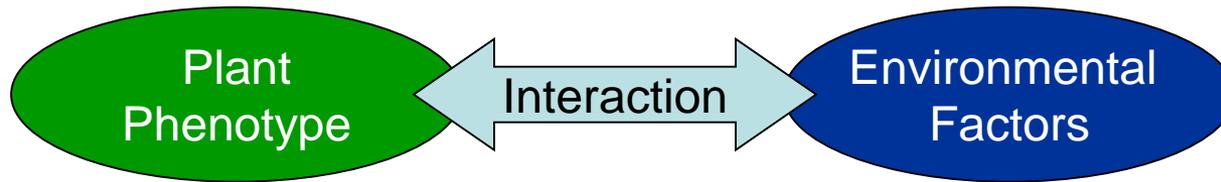
# What exactly is phenotyping?



... but soil is still a problem



# Experimental Approach



## Mechanistic understanding of the plant system by variation of

- Environmental factors: Humidity, temperature, CO<sub>2</sub>, nutrients, neighboring plants, ...
- Arabidopsis, barley, maize, tomato...
- Genotype (regulates biochemistry)

## Measurement of phenotype

- Leaf, fruit and root growth
- Photosynthesis, gas exchange, active genes, ...



PhyTec at IBG II

# An ideal Experiment

## Environment



## Genome



## Phenotype



We will image a lot, and genotype everything

Experiment time **60 days**

Sample Size **300 plants**

Genotyping **Full Sequencing**

Sequencing the plants yields **~30 TB** of data

- would take about 50 days on the HiSeq X10

Images, 3D models etc. **~300MB** per plant per day

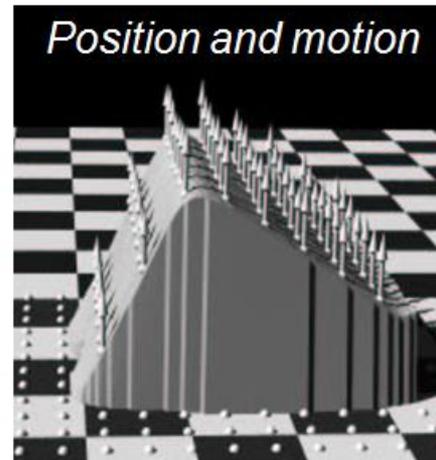
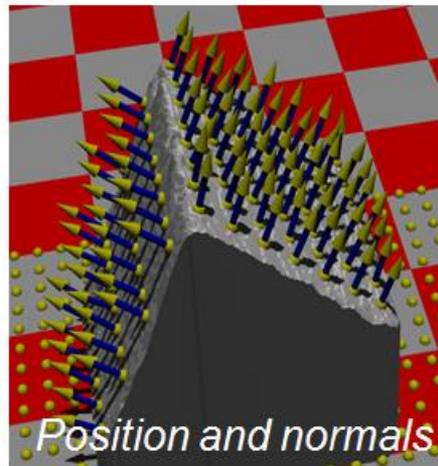
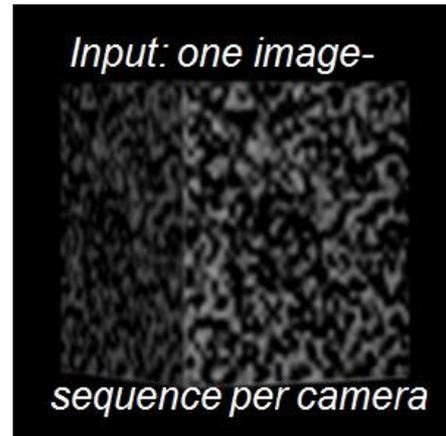
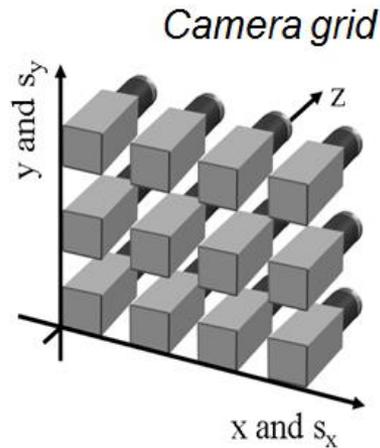
- **~6TB** per experiment

**P = GxE**: test many genotypes (species) at many environmental conditions: **big data problem**

# Image Processing @ IBG-2

## Local data

- Optical
- Transpa  
brightness
- Dynam  
reconst
- Optimiz



## Scene flow estimation from light fields

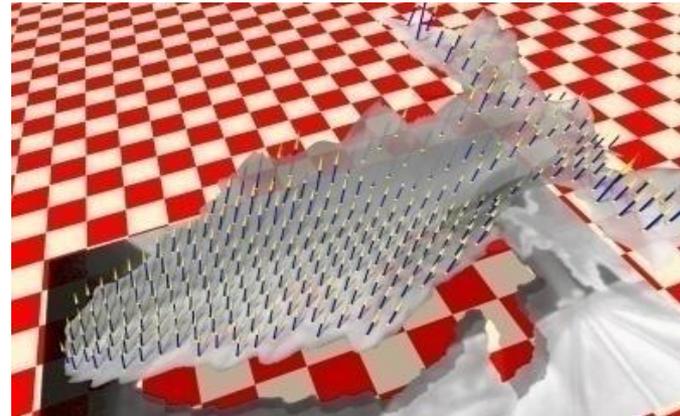
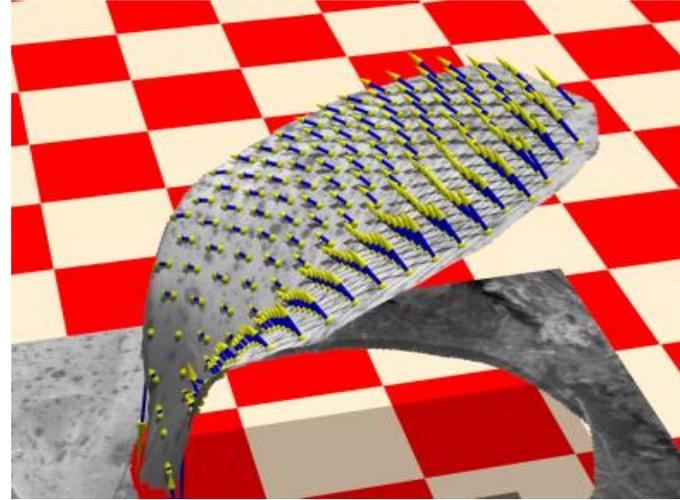
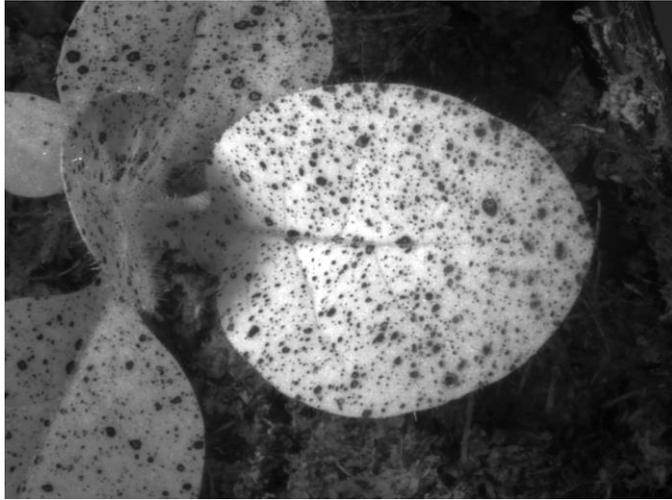
K. Krajssek, C. Heinemann, H. Scharr, Visapp 2014

T. Schuchert, T. Aach, and H. Scharr. PAMI 2010

T. Schuchert and H. Scharr. ECCV 2010

H. Scharr. Complex Motion 2007

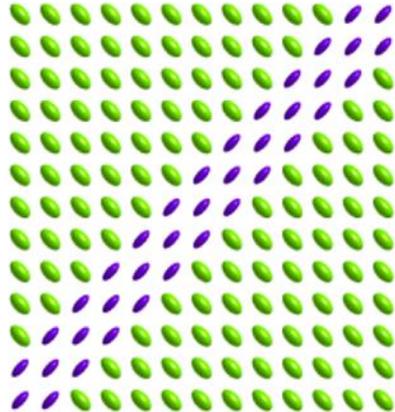
# Image Processing @ IBG-2



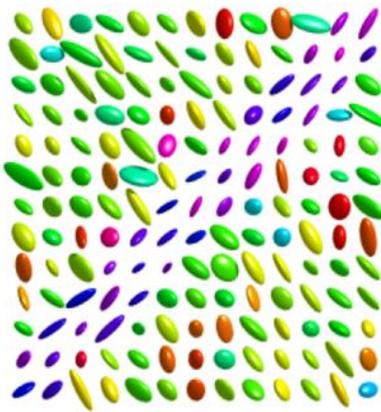
# Image Processing @ IBG-2

Local

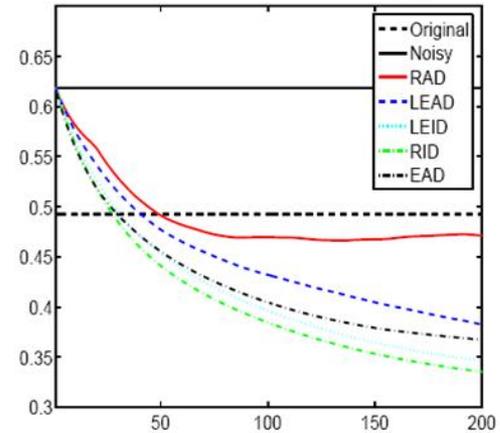
**Ground truth**



**Noise added**



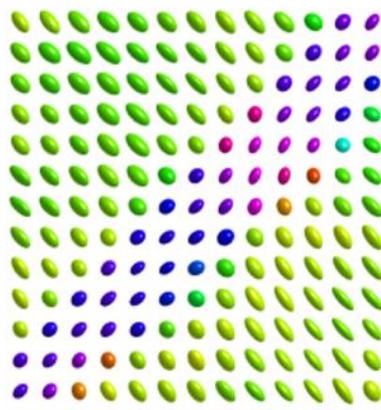
**Fractional Anisotropy**



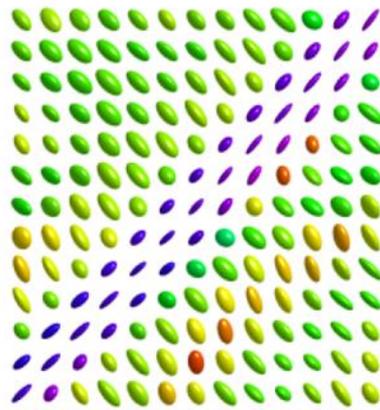
**Eukclidean flat EAD**



**Log-Eukclidean LEAD**



**Riemannian RAD**



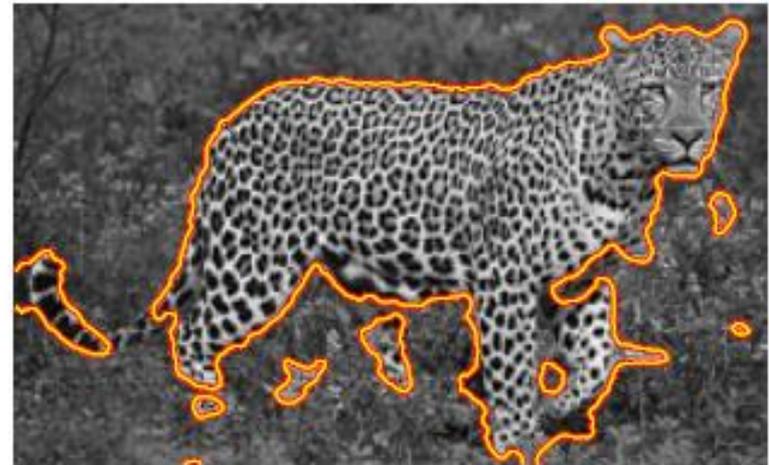
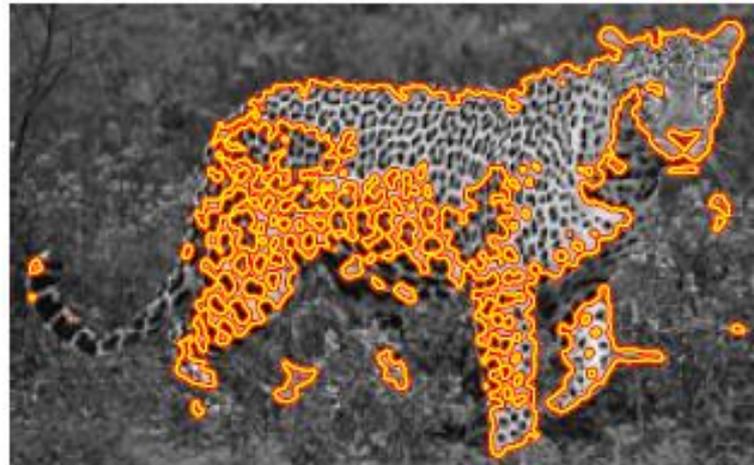
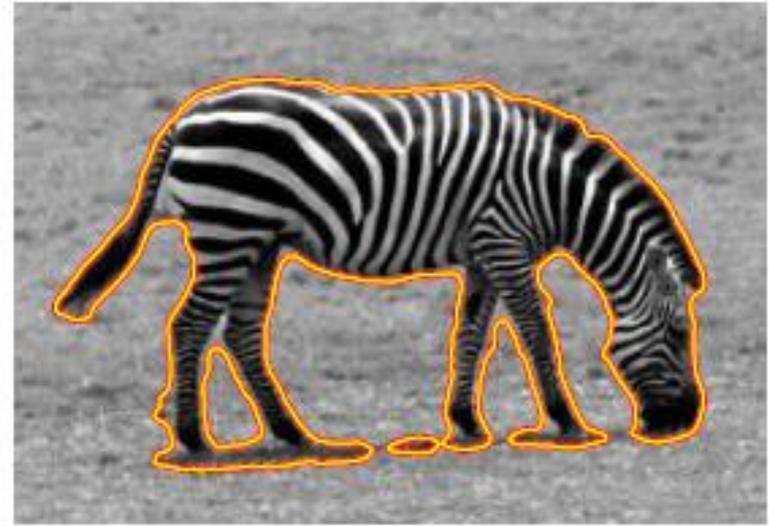
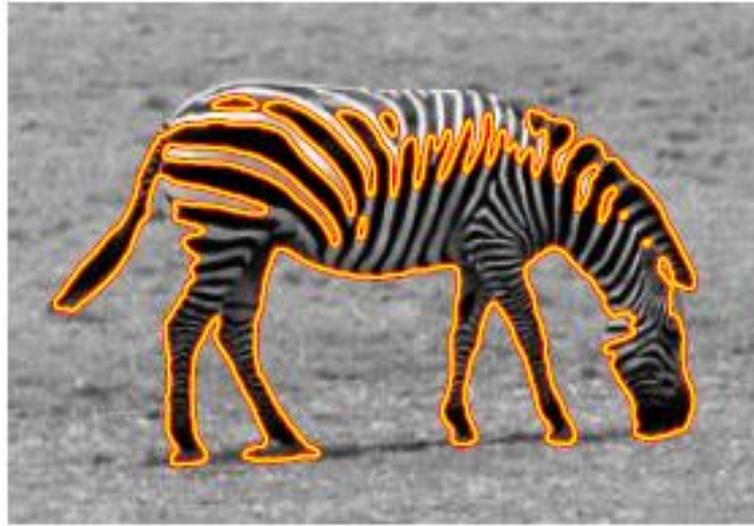
Data



Regularized Riemannian reconstruction of diffusion tensor fields from DTMRI data.

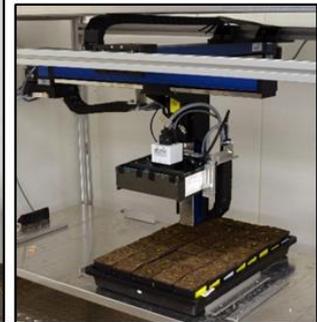
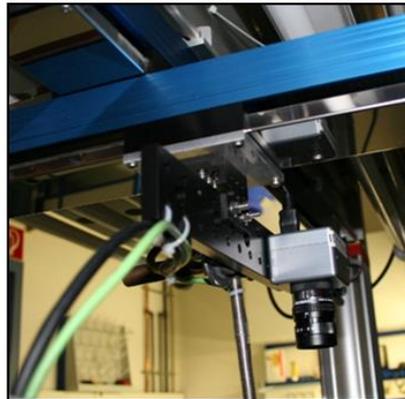
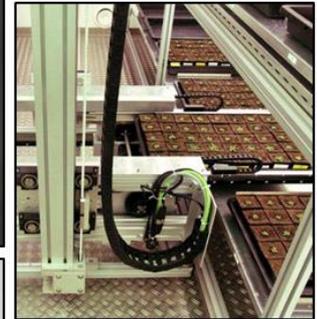
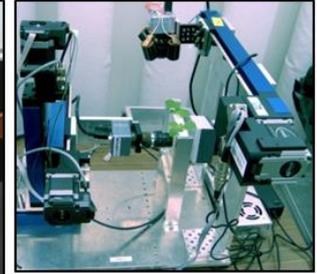
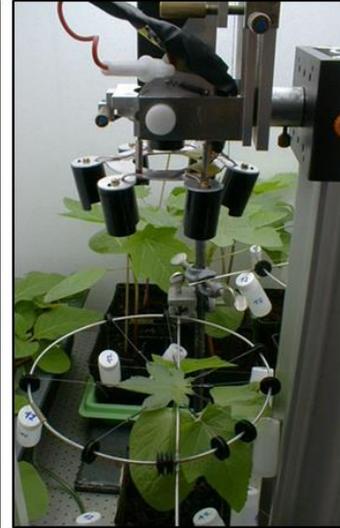
- K. Krajssek, M.I. Menzel, and H. Schar. IJCV 2015 (subm.)
- K. Krajssek and H. Schar. CVPR 2012 (HARDI)
- K. Krajssek, M.I. Menzel, and H. Schar. ICCV 2009 (DTMRI)

# Image Processing @ IBG-2



Mixture of Gaussian vs. channel representations

# Image Processing @ IBG-2



S. Bergsträsser et al., Plant Methods, 2015.  
C. Plückers et al., Nova Acta Leop. 2013.  
K.A. Nagel et al., Functional Plant Biology, 2012.  
F.-L. Luo et al., Annals of Botany, 2011.  
R.C. Meyer et al., Theoretical and Applied Genetics, 2010.  
K.A. Nagel et al., Functional Plant Biology, 2009.

M. Jansen et al., Functional Plant Biology, 2009.  
B. Biskup et al., Plant Physiology, 2009.  
R. Pieruschka et al., New Phytologist, 2008.  
A. Chavarria-Krauser et al., New Phytologist, 2008.  
A. Walter et al., New Phytologist, 2007.  
B. Biskup et al., Plant, Cell and Environment, 2007.

# Deep Phenotyping

- Precise analysis of individual traits
- Typically done at organ level

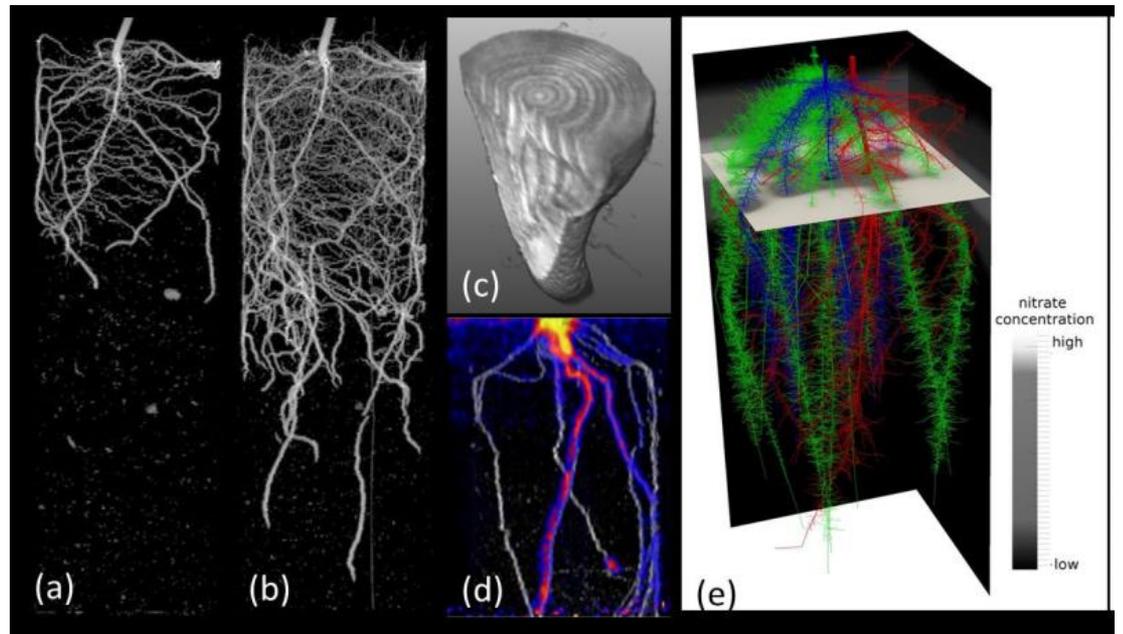
## Example: Root

### Approach :

Understand key processes of root **structure and function** for resource use efficiency



**Basis for novel root trait identification**



# MRI (Magnetic Resonance Imaging) for structural and functional imaging of roots

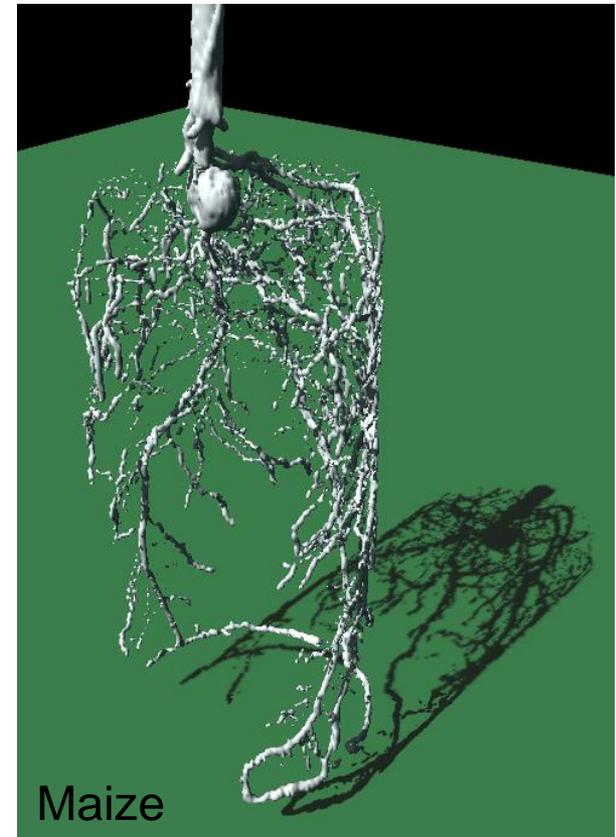


4.7T magnet, vertically oriented,  
30cm opening

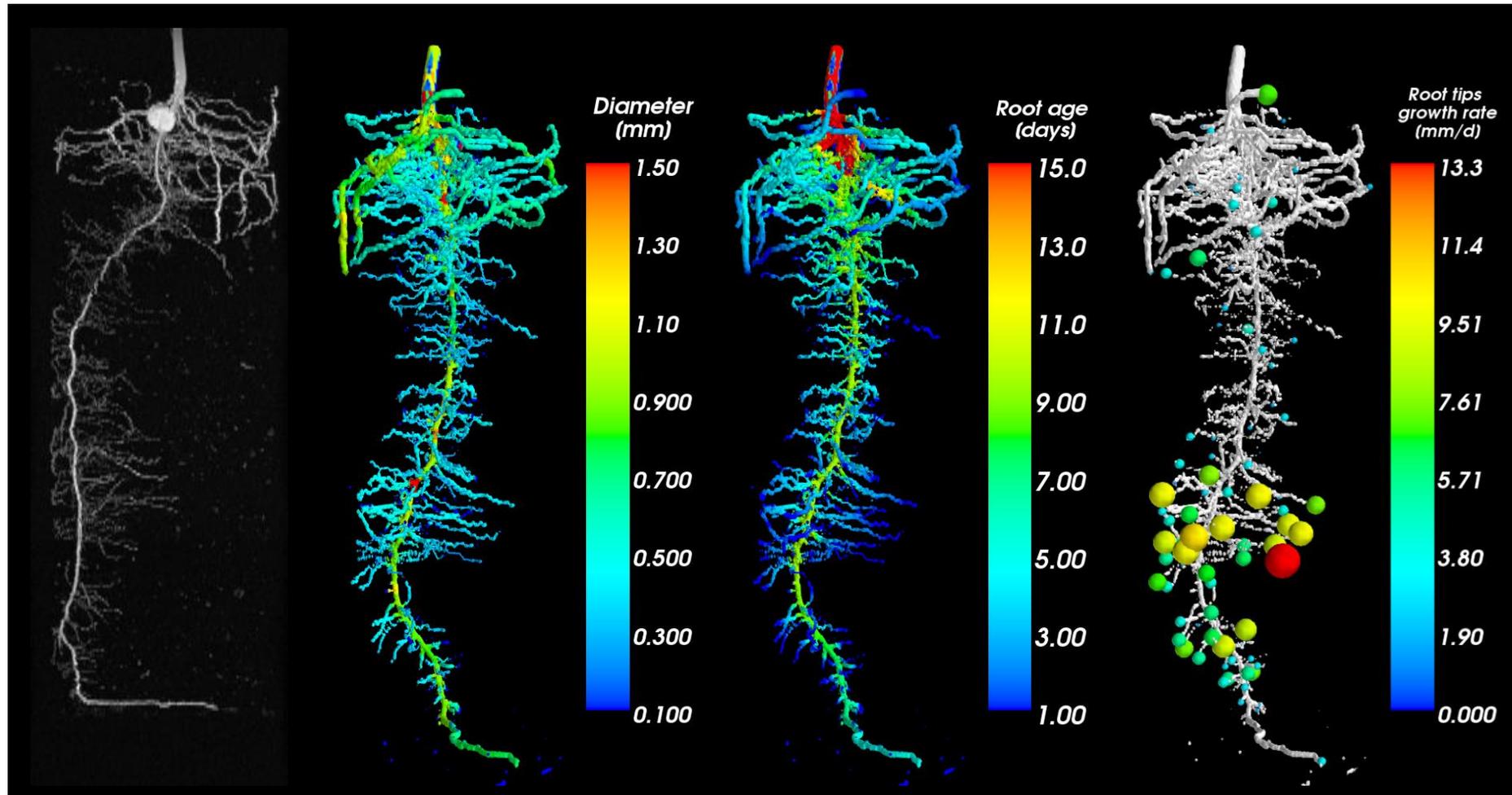


Robot systems for  
automated delivery of  
plants

**Structural analysis** of  
root system architecture  
in 3D



# Root System Analysis: 3D MRI



D. Pflugfelder, 2015, unpublished

# Screening: Seeds

- Automated measurement of seed properties
- Done individually for each single seed

## **Approach:**

Start with seed properties

Relate to plant performance  
at an early stage



**Basis for experiments with  
these plants later on**

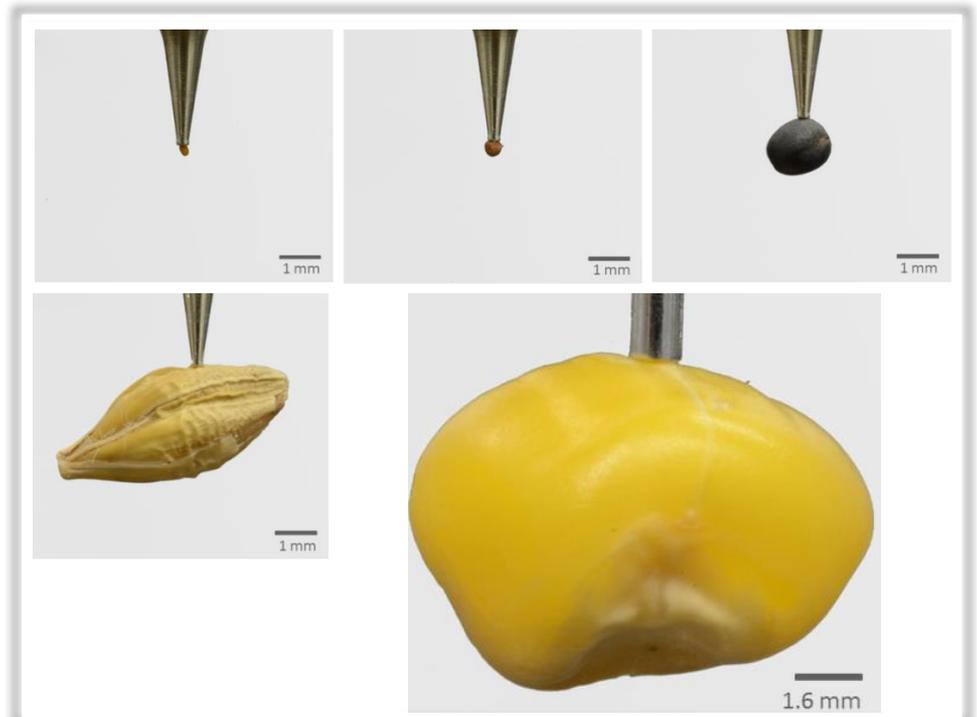
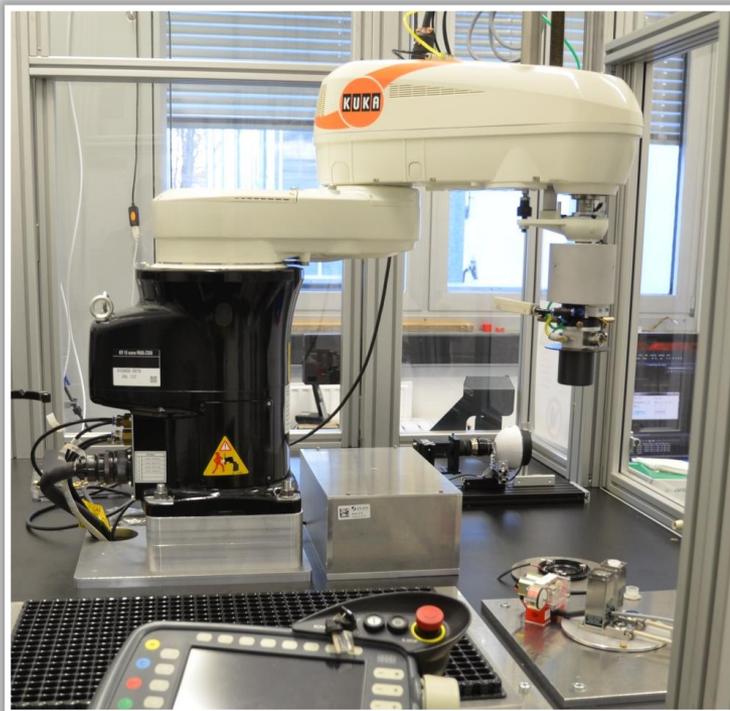




Arabidopsis seeds

# phenoSeeder

- 2D projected seed area, length, width
- Scales for weighing
- 3D shape



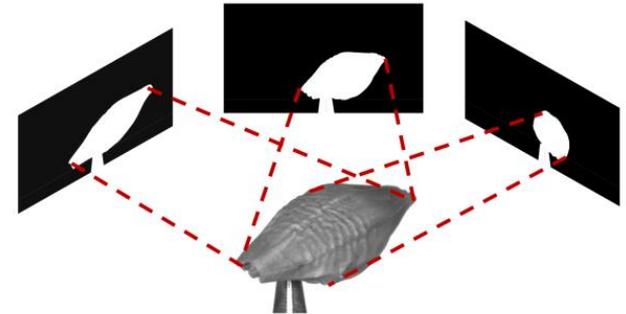
# Reconstructing Seed Shape from Silhouettes

Idea: use simple volume carving

**1. Rotate** seed in front of camera

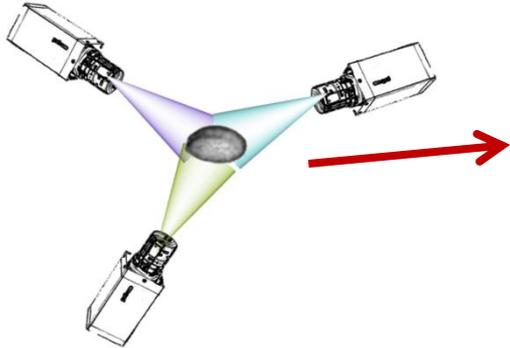


**2. Reconstruct** from **silhouettes**

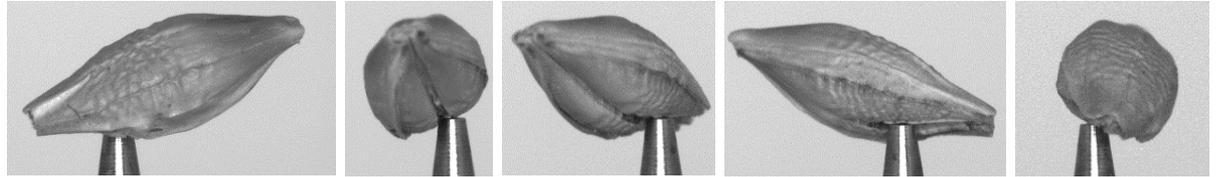


# Reconstructing Seed Shape from Silhouettes

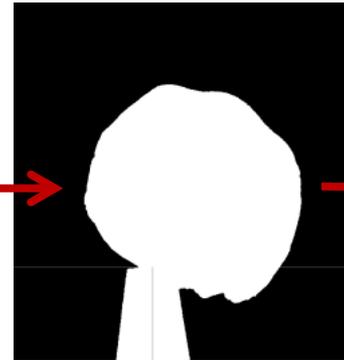
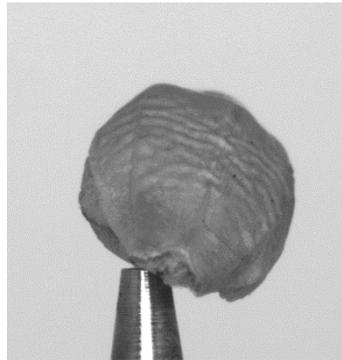
image acquisition



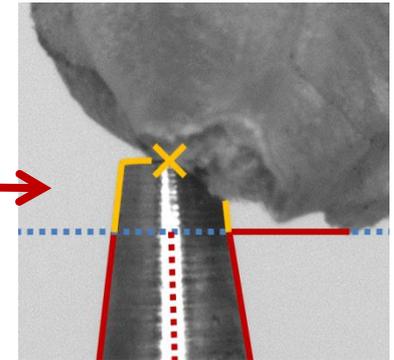
36 grey value images from different viewing angles



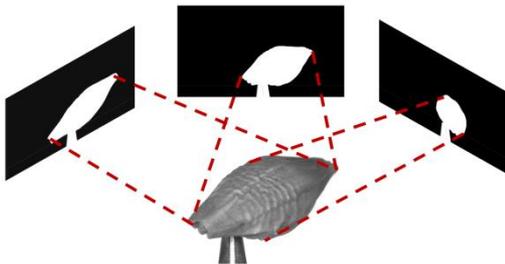
mask image



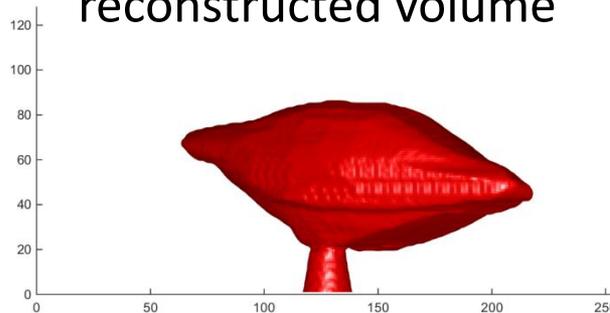
TCP detection



volume carving



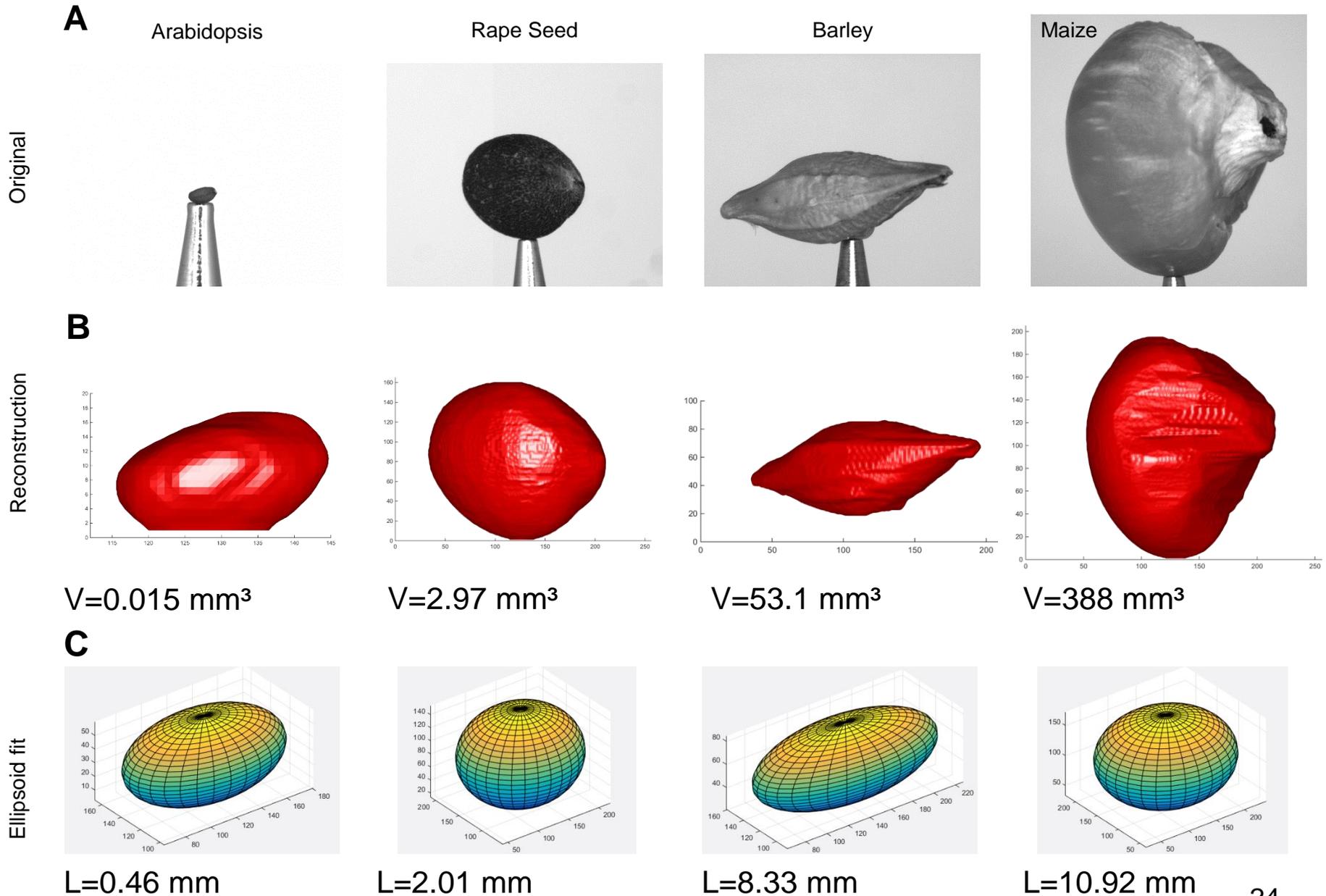
surface of reconstructed volume



tool – seed separation



# Examples – Different seeds



# Screening: Shoot

- Transfer from single plant to stand level

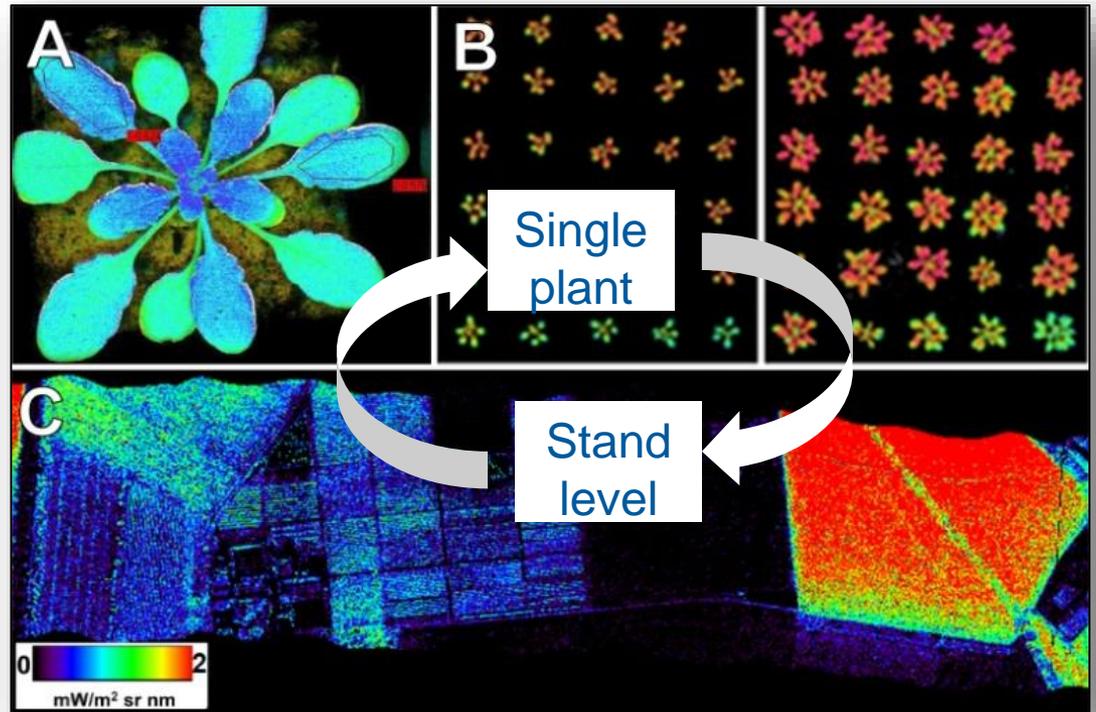
## Approach:

Dynamic environment

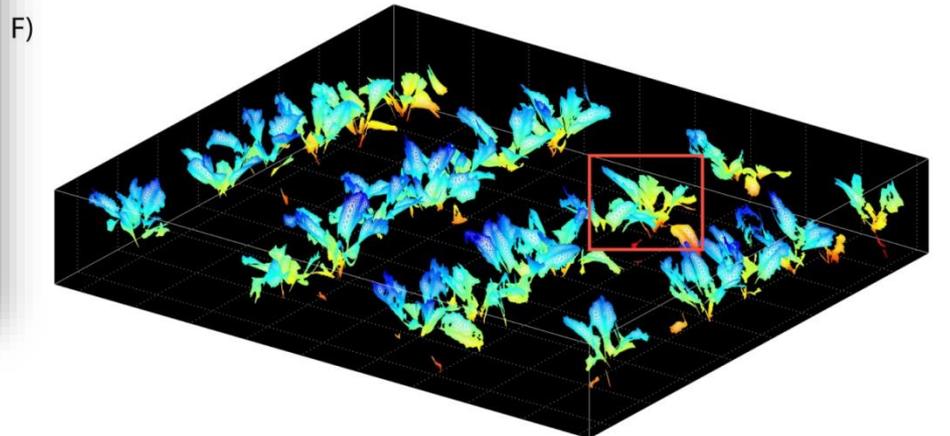
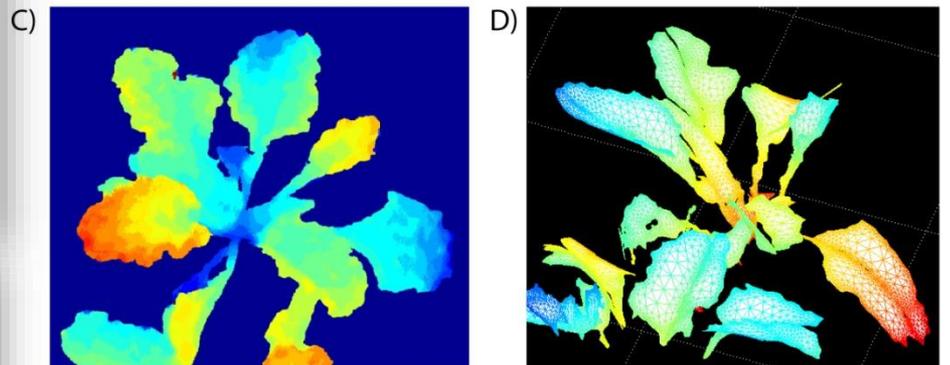
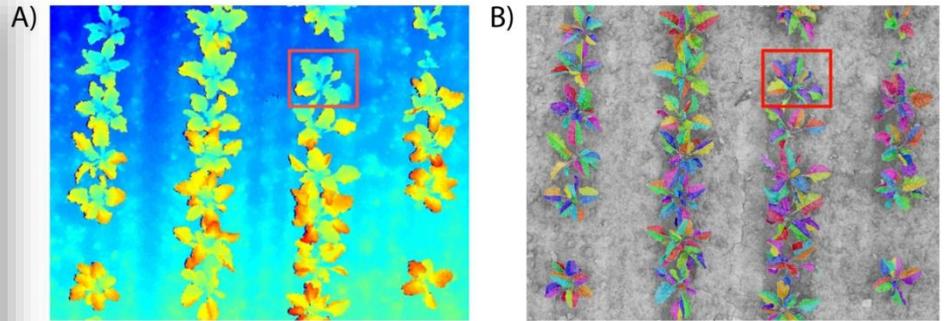
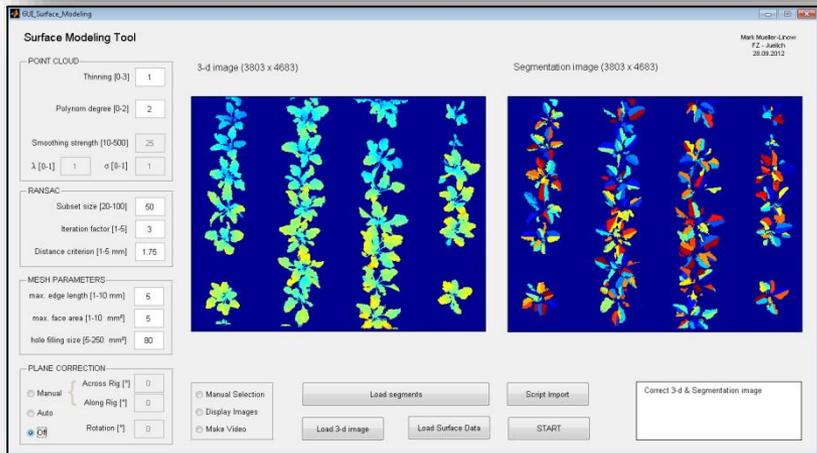
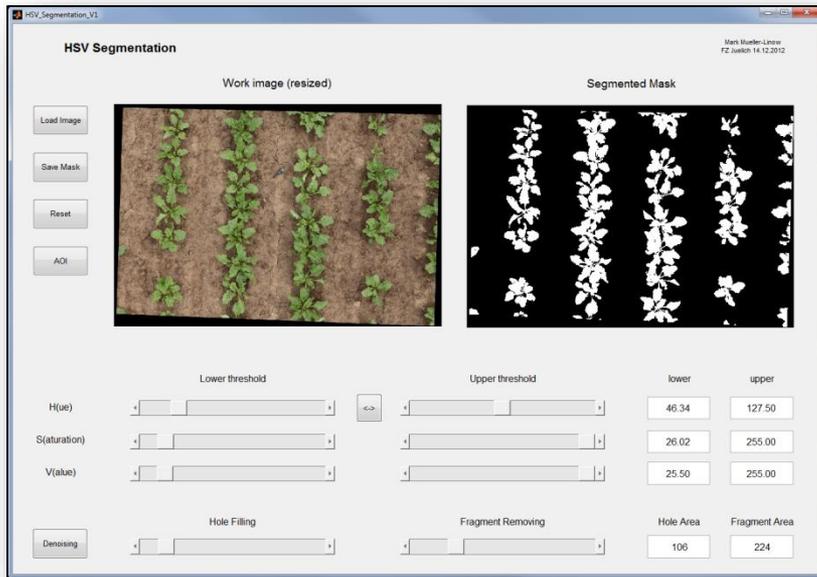
Photosynthesis and growth control from single plants to stands



**Basis for novel shoot trait identification**



# 3-D Canopy structure: Stereo imaging allows quantification of canopy structure



Müller-Linow et al., Plant Methods, 11(11), doi 10.1186/s13007-015-0052-z, 1-16, 2015

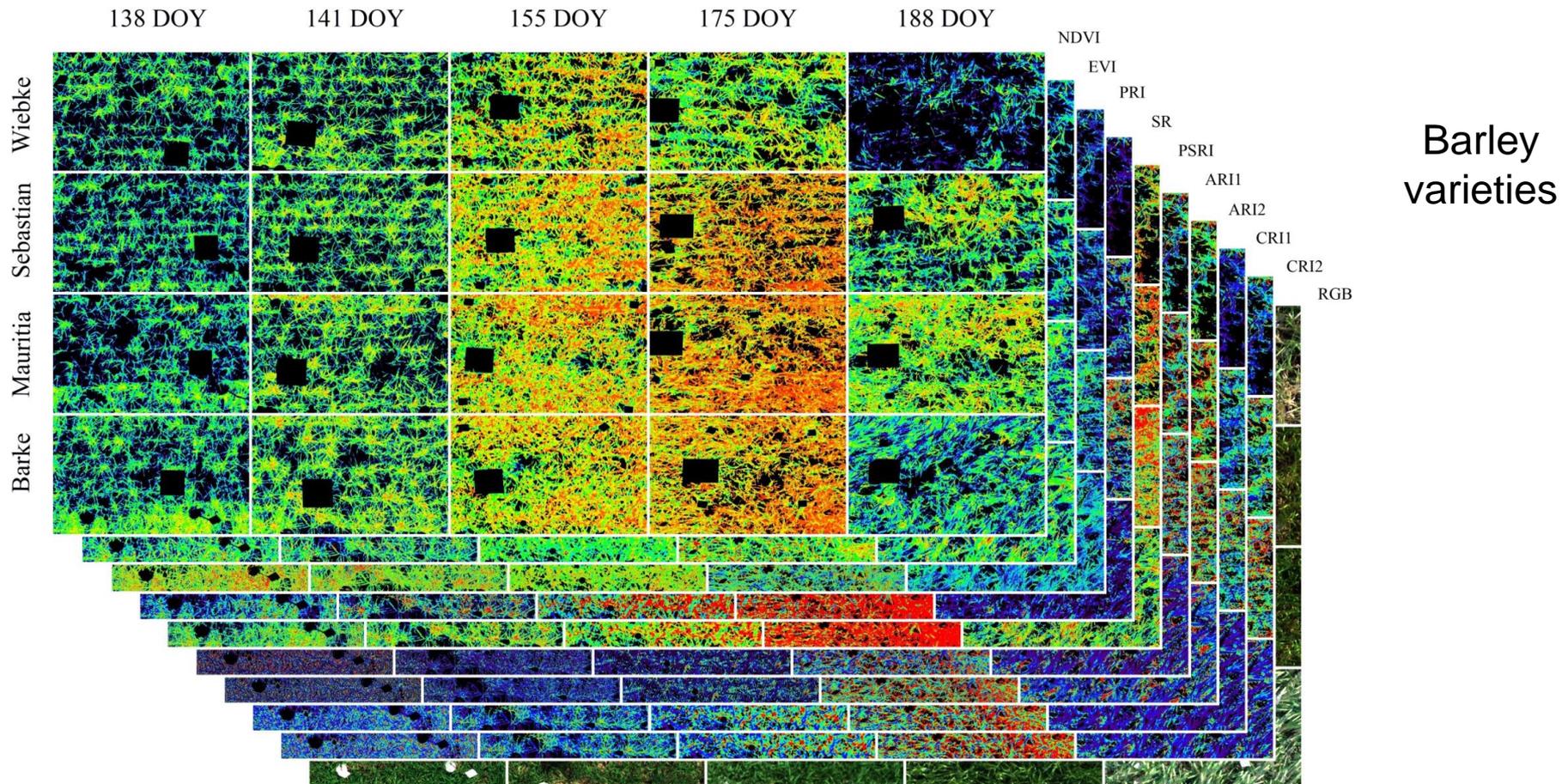
# Mapping of spatio-temporal canopy dynamics in the field by imaging spectroscopy and 3-D canopy reconstruction



Corn, sugar beet, and barley  
Measurements at 7 m  
Area: 1.5 x 2.7 m



# Mapping of spatio-temporal canopy dynamics in the field by imaging spectroscopy and 3-D canopy reconstruction



- quantify dynamics in the multidimensional data space
- relate spectral data to structural and functional aspects of canopies

# Screening: Roots

- Automated measurement of many plants
- Typically done at single plant level

## **Approach:**

High-throughput screening for root system traits

Apply heterogenous and dynamic conditions



**Screening for optimized root structure and function**

**Improve nutrient and water use efficiency**



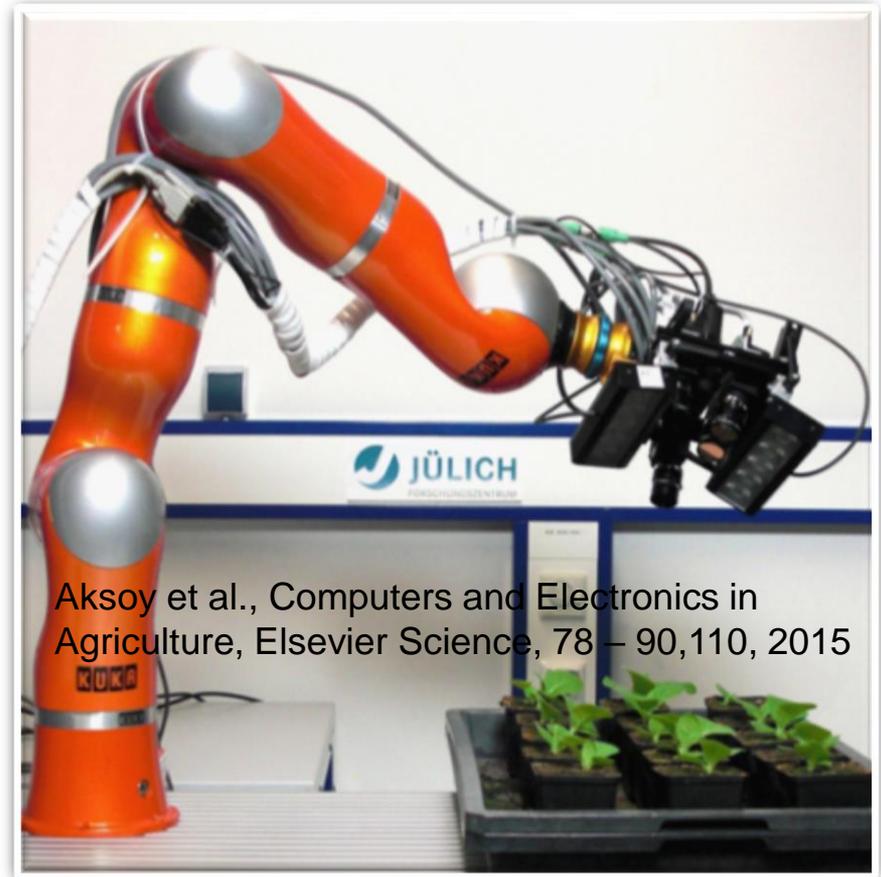
# Screening: Root System Analysis: Rhizotrons



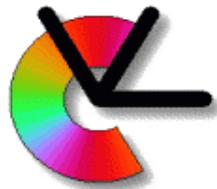
Nagel et al. *Functional Plant Biology* 2012

# GARNICS: Robot Gardener

- EU FP7 project – cognitive systems and robotics call
- GARNICS: Gardening with a Cognitive System
- Robot Gardener
  - Measure plant status from images, treatments and environmental data
  - Learn to treat plants optimally



Aksoy et al., Computers and Electronics in Agriculture, Elsevier Science, 78 – 90,110, 2015



GEORG-AUGUST-UNIVERSITÄT  
GÖTTINGEN

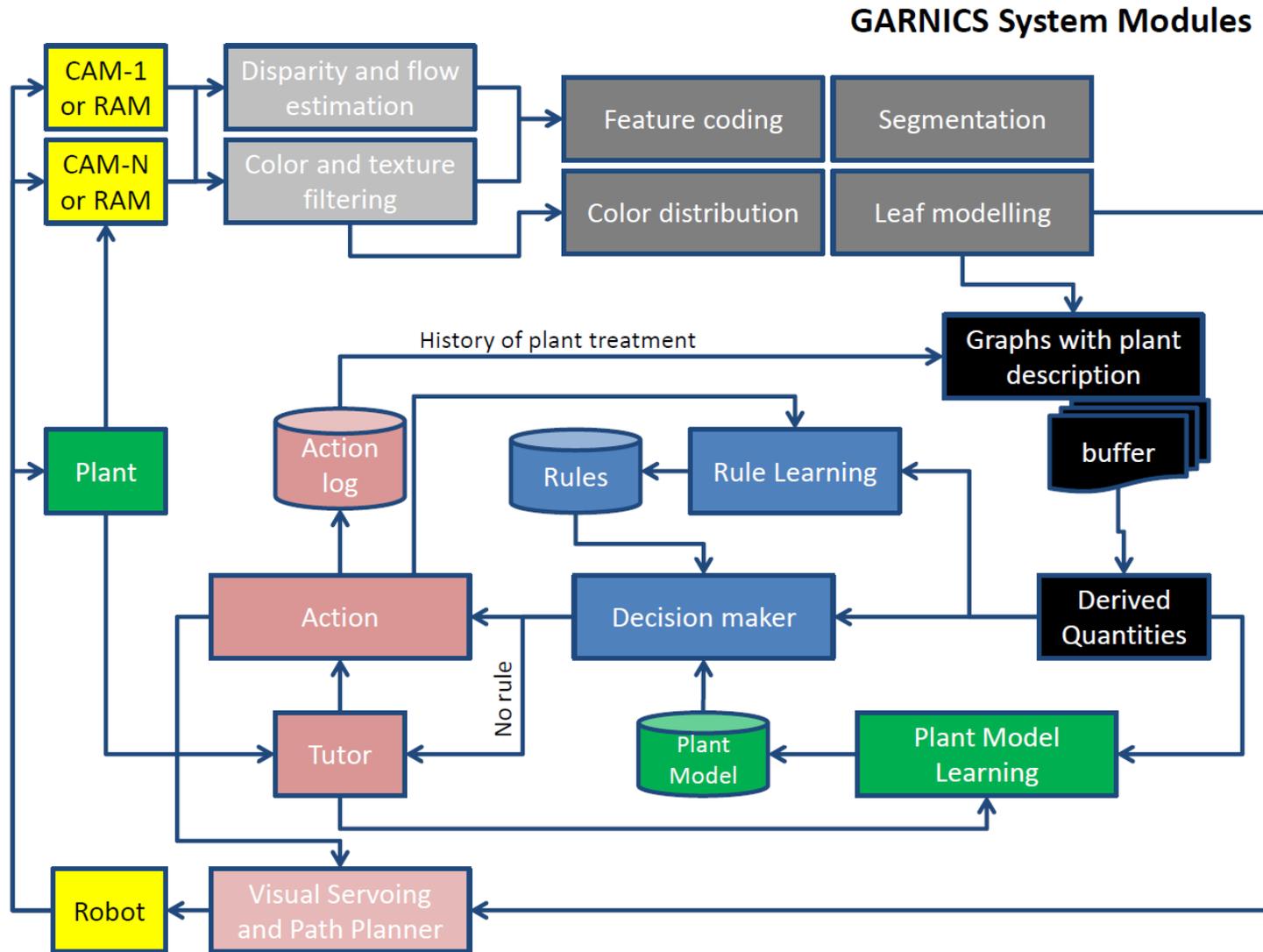


**CSIC**  
CONSEJO SUPERIOR DE INVESTIGACIONES CIENTÍFICAS

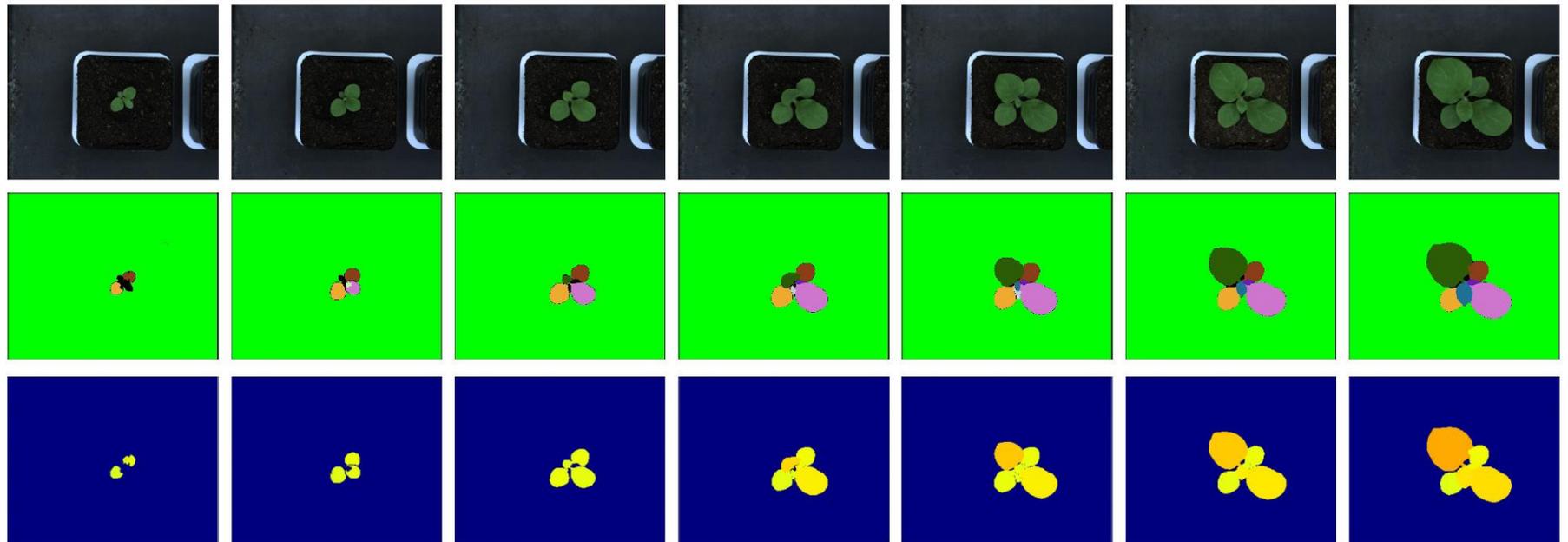
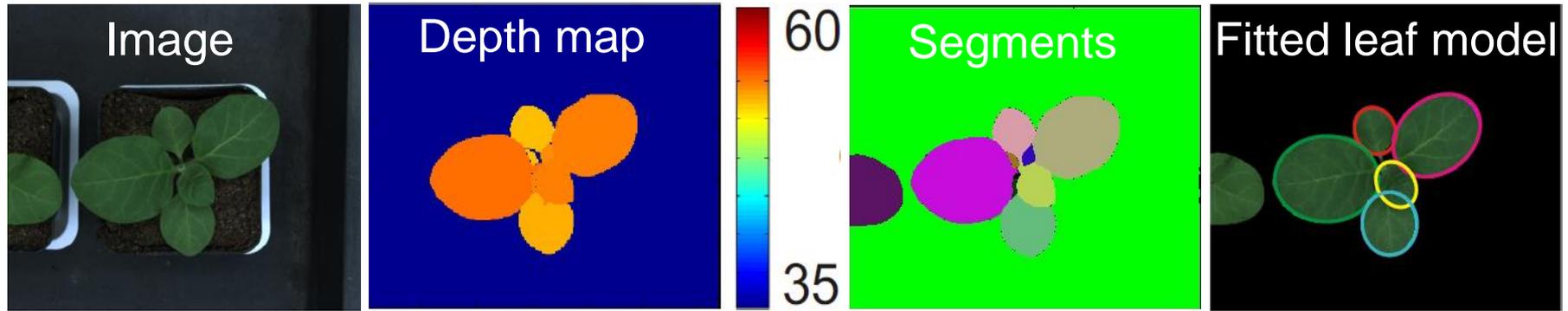


**JÜLICH**  
FORSCHUNGSZENTRUM

# Simplified GARNICS Action-Perception Loop with Memory for Plant Treatment History



# GARNICS: From Images to Plant Graphs



$$3 \cdot 2$$

$$3 \cdot \begin{matrix} 5 & 2 \\ \cdot & 4 \end{matrix}$$

$$3 \cdot \begin{matrix} 6 & 2 \\ \cdot & 4 \end{matrix}$$

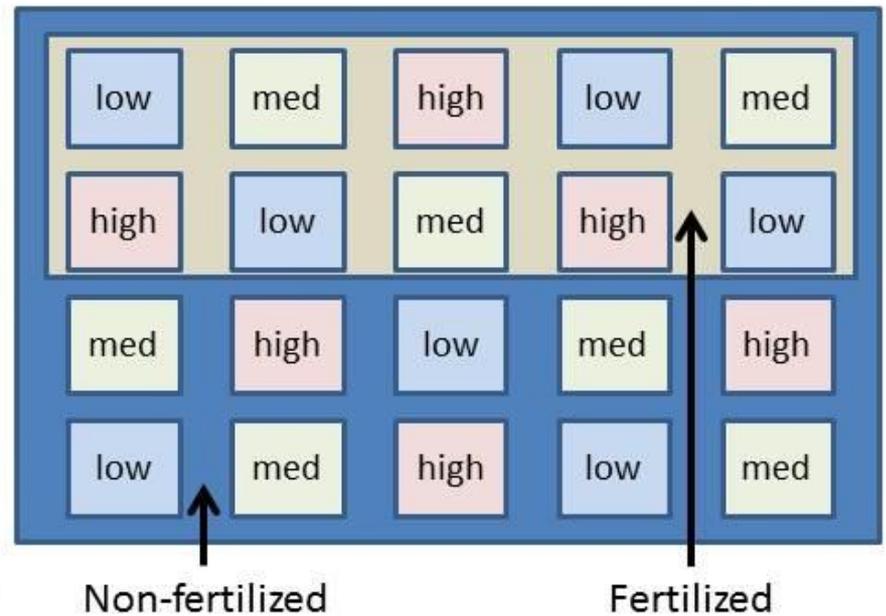
$$3 \cdot \begin{matrix} 6 & 2 & 8 \\ \cdot & 7 & 4 \end{matrix}$$

$$3 \cdot \begin{matrix} 6 & 2 \\ \cdot & 7 & 8 \\ 3 & 7 & 4 \end{matrix}$$

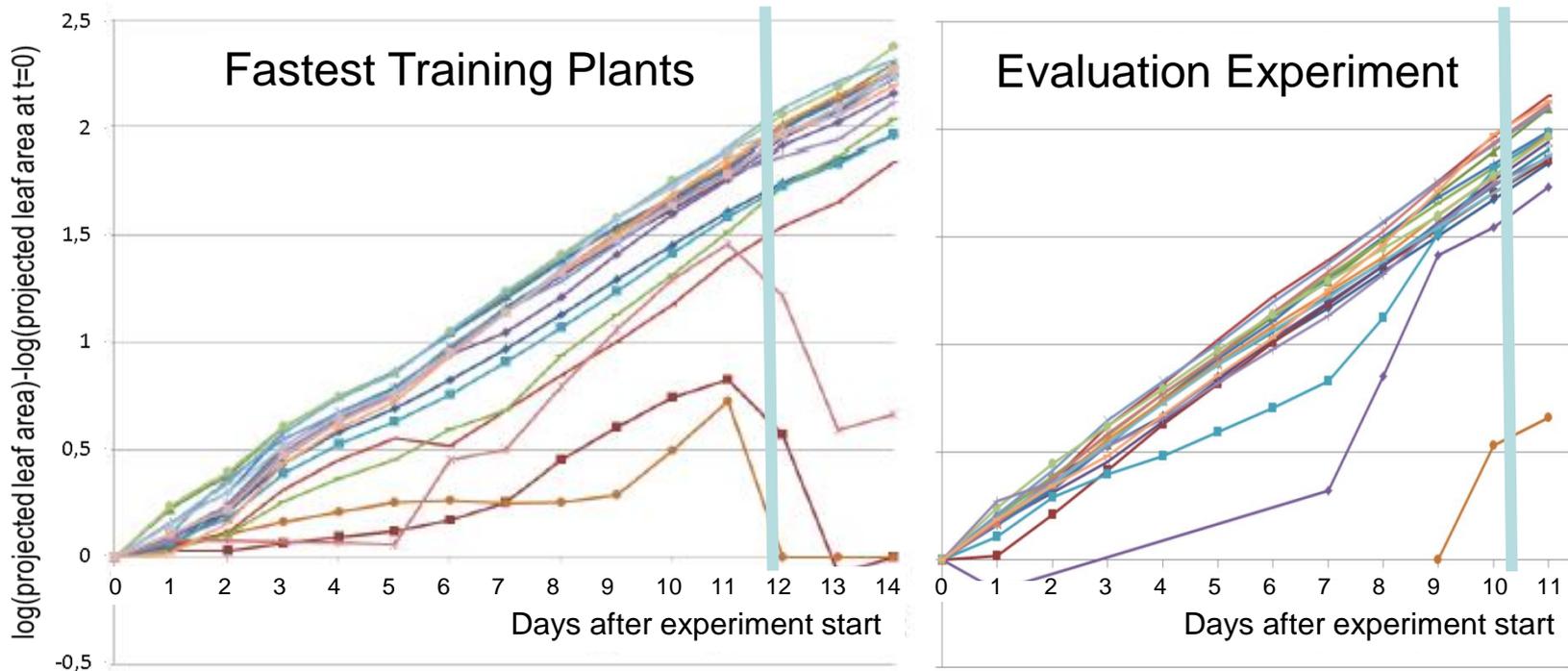
$$3 \cdot \begin{matrix} 6 & 2 \\ \cdot & 7 & 8 \\ 3 & 9 & 4 \end{matrix}$$

$$3 \cdot \begin{matrix} 6 & 2 \\ \cdot & 8 & 4 \\ 3 & 9 & 4 \end{matrix}$$

# GARNICS: Training Data

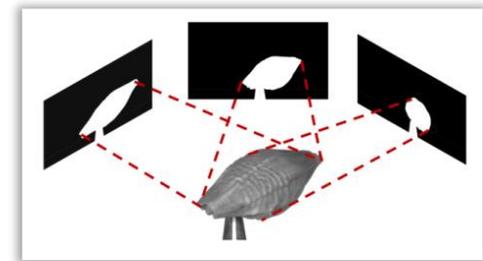
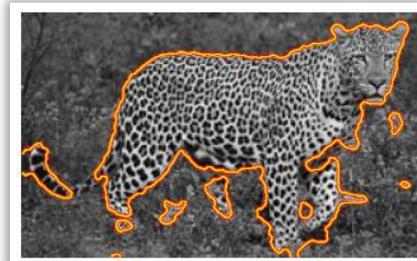
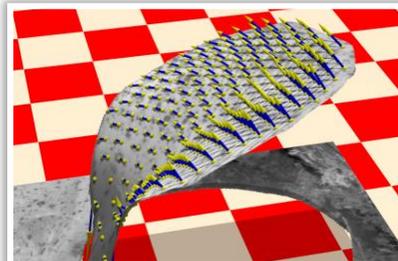
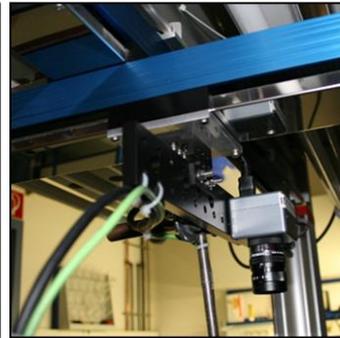
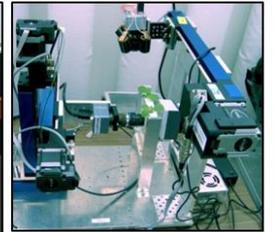
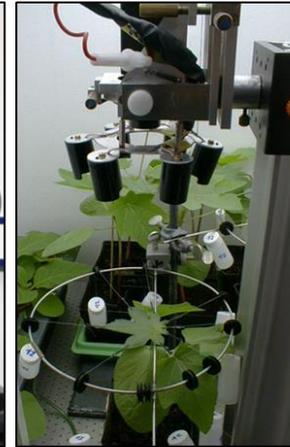
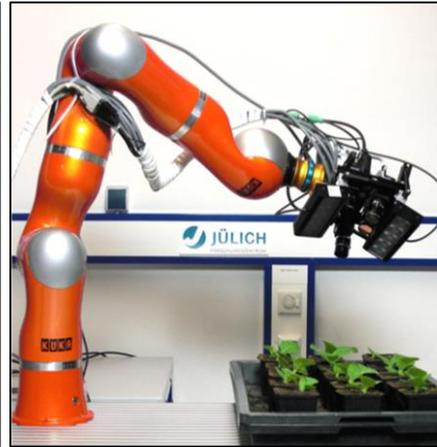
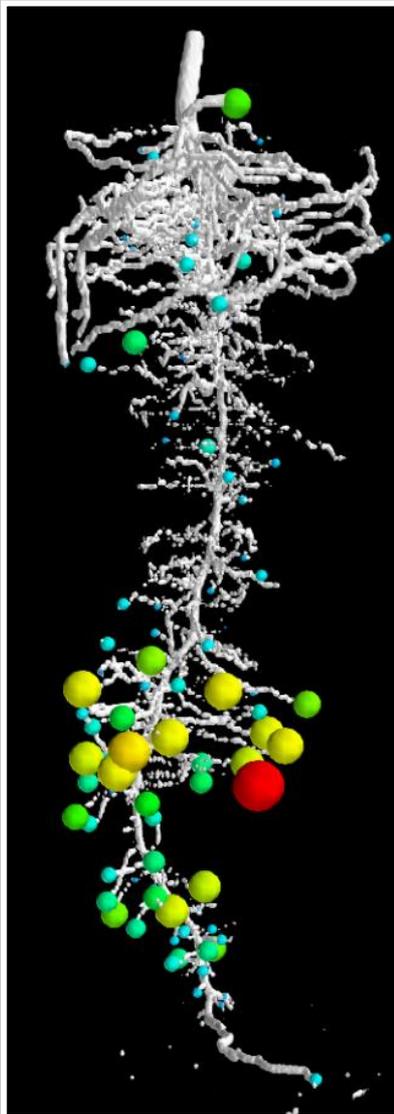


# GARNICS: Better Treatment Plan Found



- Faster growth by treatment found by the GARNICS system
- Beats best-performing training plants
- Status from day 12 reached at day 10 to 11.

# Image Processing @ IBG-2



# Imaging and Image Processing for Plant Phenotyping



